# organic papers

Acta Crystallographica Section E Structure Reports Online

ISSN 1600-5368

## K. Ravikumar,<sup>a</sup>\* B. Sridhar,<sup>a</sup> Biswanath Das<sup>b</sup> and K. Venkateswarlu<sup>b</sup>

<sup>a</sup>Laboratory of X-ray Crystallography, Indian Institute of Chemical Technology, Hyderabad 500 007, India, and <sup>b</sup>Organic Chemistry-I, Indian Institute of Chemical Technology, Hyderabad 500 007, India

Correspondence e-mail: ravikumar\_iict@yahoo.co.in

#### **Key indicators**

Single-crystal X-ray study T = 273 K Mean  $\sigma$ (C–C) = 0.003 Å R factor = 0.043 wR factor = 0.120 Data-to-parameter ratio = 10.6

For details of how these key indicators were automatically derived from the article, see http://journals.iucr.org/e.

## A clerodane diterpenoid from Pulicaria wightiana

The structure of the clerodane diterpene  $C_{21}H_{30}O_5$ , with modest antibacterial activity and isolated from the plant *Pulicaria wightiana*, was established to be methyl  $6\alpha$ -hydroxy-3,13-clerodadien-15,16-olid-18-oate. The fused six-membered rings of the decalin system have sofa and chair conformations. The furanone side chain is in an antiperiplanar conformation. The molecules in the crystal structure are stabilized by an intramolecular  $O-H\cdots O$  hydrogen bond and a chain of intermolecular  $C-H\cdots O$  interactions.

#### Comment

Various species of *Pulicaria* are distributed in different parts of India and are well known for their medicinal properties. The title compound, (I), was one of the five new clerodane diterpenoids recently isolated from the aerial part of the plant *Pulicaria wightiana*, which shows moderate activity against Gram-positive organisms, *Bacillus subtilis*, *Bacillus sphaerius* and *Staphylococeus* (Das *et al.*, 2005). A large number of diterpenoids with the clerodane skeleton have been isolated from plants in recent years (Merritt & Ley, 1992). Clerodane diterpenoids are a family of secondary metabolites found to possess considerable biological activity, *viz*. antitumoral, antimicrobial, antibacterial and particularly antifeedant (Merritt & Ley, 1992; Simmonds & Blaney, 1992). As a result of this biological potential, the present study was undertaken to obtain the three-dimensional structure of (I).



All bond lengths and angles are normal and are in good agreement with related structures reported in the literature (Spek *et al.*, 1987; Linden *et al.*, 1996; Soundarya Devi *et al.*, 2003). The molecular structure of (I) consists of a bicyclic, *viz.* cyclohexene and cyclohexane, *trans*-fused decalin ring system. It was observed that the majority of isolated clerodanes have *trans*-ring fusion (Hanson, 1999). The methyl groups at C8 and C9 are *cis* to each other  $[C21-C8-C9-C17 = 54.1 (2)^{\circ}]$ . In

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Online 4 March 2005



#### Figure 1

A view of (I), with the atom-numbering scheme. Displacement ellipsoids are drawn at the 50% probability level and H atoms are shown as small spheres of arbitrary radii.

addition, the methyl group and the hydroxy group at C5 and C6 are also *cis* to each other [C20-C5-C6-O1 = $-44.7 (2)^{\circ}$ ]. In this respect, our results support earlier spectroscopic assignments (Das et al., 2005).

As expected, the cyclohexane ring exists in a chair conformation  $[q_2 = 0.019 (2), q_3 = 0.560 (2), Q_T = 0.560 (2) \text{ Å}, \varphi_2 =$ -123 (5) and  $\theta_2 = 2.0$  (2)°; Cremer & Pople, 1975], whereas the cyclohexene ring adopts a sofa conformation with C10 lying 0.710 (2) Å out of the plane  $[q_2 = 0.416 (2), q_3 = 0.343 (2), Q_T =$ 0.540 (2) Å,  $\varphi_2 = -45.8$  (3) and  $\theta_2 = 50.6$  (3)°]. A similar conformation of the decalin ring system was observed in the crystal structure of 19-acetoxy-cis-clerodan-3-en-15-oic acid (Kolocouris et al., 2001). This conformational preference of the bicyclic decalin ring system in the present structure facilitates an intramolecular O-H···O hydrogen bonding (Table 2) between the hydroxy group at C6 and the neighboring methyl ester group at C4. The methyl ester group at C4, in an extended conformation [C19-O4-C18-C4 =177.0 (2)°], is not coplanar with the endocyclic C=C double bond  $[C3-C4-C18-O4 = -21.9 (3)^{\circ}]$ .

It is interesting to note that in some of the clerodane diterpenoids, the presence of the furan ring in the side chain appears to be one of the determinants of antifeedant activity (Enriz et al., 2000).



### Figure 2

Packing diagram viewed down the a axis. Dashed lines indicate intramolecular O-H···O hydrogen bond and intermolecular C-H...O interactions. H atoms not involved in hydrogen bonding have been omitted for clarity.

The methyl ester and furanone ring are involved in an intermolecular  $C-H \cdot \cdot \cdot O$  interaction. This links the molecules into infinite one-dimensional chains, which run parallel to the crystallographic c axis.

### **Experimental**

The title compound was isolated from the aerial parts of Pulicaria wightiana (Das et al., 2005). Needle-shaped crystals suitable for X-ray study were obtained from a methanol solution.

Crystal data

$C_{21}H_{30}O_5$	Mo $K\alpha$ radiation
$M_r = 362.45$	Cell parameters from 9796
Orthorhombic, $P_2_1 2_1 2_1$	reflections
a = 10.3572 (5) Å	$\theta = 2.2-28.0^{\circ}$
b = 11.6812 (5) Å	$\mu = 0.09 \text{ mm}^{-1}$
c = 15.5784 (7) Å	T = 273 (2) K
$V = 1884.75 (15) \text{ Å}^3$	Cut needle, colorless
Z = 4	$0.30 \times 0.14 \times 0.09 \text{ mm}$
$D_x = 1.277 \text{ Mg m}^{-3}$	

#### Data collection

Bruker SMART CCD area-detector	2386 reflections with $I > 2\sigma(I)$
diffractometer	$R_{\rm int} = 0.019$
$\omega$ scans	$\theta_{\rm max} = 28.0^{\circ}$
Absorption correction: none	$h = -13 \rightarrow 13$
16247 measured reflections	$k = -14 \rightarrow 15$
2549 independent reflections	$l = -20 \rightarrow 20$

#### Refinement

Refinement on $F^2$	$w = 1/[\sigma^2(F_0^2) + (0.0837P)^2]$
$R[F^2 > 2\sigma(F^2)] = 0.043$	+ 0.187P]
$wR(F^2) = 0.120$	where $P = (F_0^2 + 2F_c^2)/3$
S = 1.04	$(\Delta/\sigma)_{\rm max} < 0.001$
2549 reflections	$\Delta \rho_{\rm max} = 0.28 \text{ e } \text{\AA}^{-3}$
240 parameters	$\Delta \rho_{\rm min} = -0.18 \text{ e } \text{\AA}^{-3}$
H-atom parameters constrained	

### Table 1

Selected geometric parameters (Å, °).

O2-C15	1.199 (3)	C3-C4	1.332 (3)
O3-C15	1.365 (4)	C13-C14	1.324 (3)
O5-C18	1.214 (2)		
C4-C5-C10	105.34 (13)	C7-C8-C9	110.86 (15)
C7-C6-C5	112.95 (14)	C8-C9-C10	107.19 (13)

Table 2	
Hydrogen-bond geometry (Å, $^{\circ}$ ).	

$\overline{D-\mathrm{H}\cdots A}$	$D-\mathrm{H}$	$H \cdot \cdot \cdot A$	$D \cdots A$	$D - H \cdots A$
$O1-H1\cdots O5$ $C20-H204\cdots O5$	0.82	1.87	2.662(2)	164 126
$C19-H19C\cdots O2^{i}$	0.96	2.50	3.162 (3)	120

Symmetry code: (i) x, y, z + 1.

The absolute configuration could not be established in this analysis and was assigned on the basis of the spectroscopically found configuration of the title compound (Das *et al.*, 2005). In the absence of anomalous scattering effects, Friedel pairs were merged. H atoms were included in calculated positions (C–H = 0.93–0.98 Å) using a riding model, with  $U_{\rm iso}$ (H) values set at 1.2 (O and CH H atoms) and 1.5 (CH3) times the  $U_{\rm eq}$  values of the parent atoms.

Data collection: *SMART* (Bruker, 2001); cell refinement: *SAINT* (Bruker, 2001); data reduction: *SAINT*; program(s) used to solve structure: *SHELXS97* (Sheldrick, 1997); program(s) used to refine structure: *SHELXL97* (Sheldrick, 1997); molecular graphics: *SHELXTL/PC* (Sheldrick, 1990) and *PLATON* (Spek, 2003); software used to prepare material for publication: *SHELXL97* and *PARST* (Nardelli, 1995).

The authors thank Dr J. S. Yadav, Director of IICT, for his kind encouragement and support.

### References

- Bruker (2001). SAINT (Version 6.28a) and SMART (Version 5.625). Bruker AXS Inc., Madison, Wisconsin, USA.
- Cremer, D. & Pople, J. M. (1975). J. Am. Chem. Soc. 97, 1354-1358.
- Das, B., Ravinder Reddy, M., Ramu, R., Ravindranath, N., Harish, H., Ramakrishna, K. V. S., Koteswar Rao, Y., Harakishore, K. & Murty, U. S. N. (2005). *Phytochemistry*. In the press.
- Enriz, R. D., Baldoni, H. A., Zamora, M. A., Jauregui, E. A., Sosa, M. E., Tonn, C., Luco, J. M. & Gordaliza, M. (2000). *J. Agric. Food Chem.* **48**, 1384–1392.
- Hanson, J. R. (1999). Nat. Prod. Rep. 16, 209–219.
- Kolocouris, A., Mavromoustakos, T., Demetzos, C., Terzis, A. & Grdadolnik, S. G. (2001). Bioorg. Med. Chem. Lett. 11, 837–840.
- Linden, A., Juch, M. & Ruedi, P. (1996). Acta Cryst. C52, 933-935.
- Merritt, A. T. & Ley, S. V. (1992). Nat. Prod. Rep. 9, 243-287.
- Nardelli, M. (1995). J. Appl. Cryst. 28, 659-1142.
- Sheldrick, G. M. (1990). SHELXTL/PC. Bruker AXS Inc., Madison, Wisconsin, USA.
- Sheldrick, G. M. (1997). SHELXS97 and SHELXL97. University of Göttingen, Germany.
- Simmonds, M. S. J. & Blaney, W. M. (1992). Advances in Labiatae Science, edited by R. M. Harley & T. Reynolds, pp. 375–392. Kew, England: Royal Botanic Gardens.
- Soundarya Devi, S., Malathi, R., Rajan, S. S., Aravind, S., Krishnakumari, G. N. & Ravikumar, K. (2003). *Acta Cryst.* C**59**, 0530–0532.
- Spek, A. L. (2003). J. Appl. Cryst. 36, 7-13.
- Spek, A. L., Duisenberg, A. J. M., Labadie, R. P., Ratnayake, S., Abeysekera, A. & De Silva, K. T. D. (1987). Acta Cryst. C43, 530–532.